

**Athens Institute for Education and Research
ATINER**



**ATINER's Conference Paper Series
AGR2015-1642**

**Effects of Proline Applications on Yield and
Quality Parameters in Kapija Pepper Grown
under Different Irrigation Levels-2**

**Tolga Sariyer
Research Assistant
Canakkale 18 Mart University
Turkey**

**Canan Oztokat Kuzucu
Canakkale 18 Mart University
Turkey**

An Introduction to
ATINER's Conference Paper Series

ATINER started to publish this conference papers series in 2012. It includes only the papers submitted for publication after they were presented at one of the conferences organized by our Institute every year. This paper has been peer reviewed by at least two academic members of ATINER.

Dr. Gregory T. Papanikos
President
Athens Institute for Education and Research

This paper should be cited as follows:

Sariyer, T. and Oztokat Kuzucu, C. (2015). "Effects of Proline Applications on Yield and Quality Parameters in Kapija Pepper Grown under Different Irrigation Levels-2", Athens: ATINER'S Conference Paper Series, No: AGR2015-1642.

Athens Institute for Education and Research
8 Valaoritou Street, Kolonaki, 10671 Athens, Greece
Tel: + 30 210 3634210 Fax: + 30 210 3634209 Email: info@atiner.gr URL:
www.atiner.gr

URL Conference Papers Series: www.atiner.gr/papers.htm

Printed in Athens, Greece by the Athens Institute for Education and Research. All rights reserved. Reproduction is allowed for non-commercial purposes if the source is fully acknowledged.

ISSN: 2241-2891

13/10/2015

Effects of Proline Applications on Yield and Quality Parameters in Kapija Pepper Grown Under Different Irrigation Levels-2

Tolga Sariyer
Research Assistant
Çanakkale 18 Mart University
Turkey

Canan Oztokat Kuzucu
Çanakkale 18 Mart University
Turkey

Abstract

Turkey is among the countries at risk in terms of global warming, therefore precautions for drought are necessary. Effects of drought stress to the fatty acid composition of the kapija pepper is unknown. This research was done at the research and practice field of Çanakkale 18 Mart University to determine the effects of different irrigation levels and proline applications on yield, chlorophyll and some fatty acid compositions of pepper (*Capsicum annum* var. *conoides* Mill.) cv. Yalova Yağlık-28 variety. This experiment was laid out in a factorial setted randomized block design with 3 replications. Plants were irrigated with 3 different irrigation levels (Kcp1: 0.5, Kcp2: 1, Kcp3: 1,5) and also proline was applied to all subjects except the control group. Linoleic and alpha linolenic acids were found to be major acids and the effects of deficit irrigation and proline applications on the fatty acid composition of Yalova Yağlık-28 (*Capsicum annum* L.) were found statistically significant.

Keywords: *Capsicum annum* L., Irrigation level, Proline

Introduction

Pepper is a cultivated plant which grows annual in warm climates and perennial in tropical climates. Bailey, classifies the peppers as cherry peppers, conical peppers, red fascicular peppers, long sharp peppers and bell peppers and mentions the need for optimal water request and that the roots are sensitive to excess amounts of water. Therefore the amount of water that peppers need have to be given without interruption (Vural, 2000).

Pepper (*Capsicum annuum* L.) is one of the most susceptible crops to drought stress and moderately susceptible to salt stress (Alvino et al., 1994; Ayers and Westcot, 1989; Meiri and Shalhevet, 1973; Rhoades et al., 1992).

From the terrains classified according to stress factors, drought stress has the highest percent (%26) followed by the mineral stress (%20) and cold stress (%15). Other stress factors have 29 percent (Blum, 1986).

In water deficit conditions closed stomata can't allow the intake of CO₂ and electrons which have to take part in the reduction of CO₂, interacting with O₂ and create 'Active O₂ Radicals' like superoxide (O⁻) (Oztekin, 2009).

Five drought sensitive and drought resistant chickpea cultivars studied to determine proline accumulation, protein profile and DNA polymorphism in water deficit conditions by Ahire et al. (2005). Consequently drought resistant chickpea cultivars had a higher proline accumulation than drought sensitive cultivars.

In plant tissues, the most abundant saturated fatty acids are palmitic and stearic, and the most common unsaturated fatty acids are oleic, linoleic, and linolenic (Murphy, 1993).

Fatty acids which have one or more double bonds between carbon atoms are known as poly unsaturated fatty acids. The most important of these fatty acids are Linoleic acid; [C18:2 (n-6 omega)], α -linolenic acid; [C18:3 (n-3 omega)], arachidonic acid; [C20:4 (n-6 omega)], Eikosapentaenoic acid; [C20:5 (n-3 omega)] and Dokosahekzaenoic acid; [C22:6 (n-3 omega)] (Gogus and Smith, 2010; Holub, 2002).

α -linolenic acid (C18:3) is part of the n-3 series, and it is known to be present in many plants (Ghafoorunissa & Pangrekar, 1993; Pereira *et al.*, 2001; Liu *et al.*, 2002).

Perez-Galvez et. al. (1999) determined that fatty acid compositions in the pericarp of two pepper varieties (*Capsicum annuum* L. cv. Jaranda and Jariza) were; linolenic (29.93%), linoleic (27.15%), palmitic (18.22%), oleic (8.48%), myristic (7.16%) and stearic (4.73%), from major to minor respectively on Jaranda and linolenic (30.27%), linoleic (25.18%), palmitic (18.72%), myristic (8.27%), oleic (7.19%) and stearic (4.90%) on Jariza variety.

Fatty acid compositions were approximate in three pepper varieties (Arnoia, Fresno de la and Los Valles-Benavente) and in three ripening stages. Linoleic acid was the most abundant fatty acid followed by linolenic and palmitic acids which contain more than 80 percent of total fatty acids (Martinez et al., 2006).

Drought stressed *Arabidopsis thaliana* (Ecotype Columbia) leaves represented 61% of total fatty acid content at a 47.5% relative water content.

Polar lipid contents and lipid composition of *Arabidopsis thaliana* leaf membranes were stable under water stress with an enhance in fatty acid unsaturation (Gigon et al., 2004).

Drought tolerant plants reduce the effects of stress through cellular mechanisms, however drought sensitive plants undergo irrecoverable cell damage (Vieira da Silva et al., 1974).

Rape plants (*Brassica napus*) decreased linolenic acids mainly in chloroplast monogalactosyldiacylglycerol (MGDG) and decreased linoleic acids in phospholipid fractions in drought stress (Dakhma et al., 1995).

Drought tolerant (Tifway) and drought sensitive (C299) bermudagrass genotypes were grown under control and drought conditions. Unsaturated fatty acids, particularly linoleic, linolenic reduced and palmitic, stearic acids increased in both genotypes under water stress conditions while Tifway had more unsaturated fatty acids, mainly linoleic acids and less stearic and palmitic acids (Huang et al., 2011).

This research has been laid out in order to determine the yield and some quality parameters of the red pepper (*Capsicum annuum* var. *conoides* Mill.), an important export product of Turkey-Çanakkale which had been planted in different irrigation regimes and in an exogenous proline application.

Materials and Methods

This research was conducted at the research and practice field of Çanakkale 18 Mart University. The fertilization program was implemented according to soil analysis and results have been taken from 0-20 cm depth as represented in the terrain before the beginning of the trial. Seedlings of the Yalova Yaglik-28, Kapija pepper (*Capsicum annuum* L.) were used as plant material.

Irrigations were performed by the measuring of evaporation from a class a pan evaporation container and by applying three different pan coefficients (Kcp1: 0.5, Kcp2: 1, Kcp3: 1.5) with drip irrigation and constant pressure lateral pipes (4 liters/hour) (Yildirim, 1996).

The plants were divided into three groups for exogenous proline applications and sprayed after the 30 days from the seedling planting date. 12 mM proline applications were sprayed to all plants by spraying them from the upper and lower parts of leaves.

This experiment was laid out on a randomized block split plot design with 3 replications, consisting of 5 rows and 7 plants in each row and a total of 35 plants for each subject. The research was completed with (35*6*3) 630 plants and 18 parcels. The 2 outer rows on each side were left as edge effect and the analysis and measurements were performed in 10 randomly selected plants for each replication.

Subjects of the research:

1. Kcp1:0.5, No Proline Application
2. Kcp1:0.5, 12mM Proline (30th day from seedling planting)
3. Kcp2:1, No Proline Application
4. Kcp2:1, 12mM Proline (30th day from seedling planting)
5. Kcp3:1.5, No Proline Application
6. Kcp3:1.5, 12mM Proline (30th day from seedling planting)

Physical And Chemical Analysis

Fresh Fruit Weight (g)

The fruit weight (g) was determined by measuring the scales and the means calculated, with precision.

Yield per Plant (g/plant)

The yield per plant (g/plant) was determined by collecting all the fruit weights from yield plants when using 0.33 of meter row and 1.20 meter inter row spacing.

Fruit Number

The fruit number determined by counting the fruits and means calculated.

Total Chlorophyll ($\mu\text{g}/100\text{cm}^2$)

The chlorophyll amount in the leafs was determined with the spectrometric method (Holden, 1976).

Non-diseased, matured and homogenous 6th leafs were collected from the bottom of all applications. The leafs were carried to the laboratory with containers. 4 g of discs cut out from the leafs were extracted in %90 acetone and a solution distilled by wattman no 2 filter paper. The solution was used for the absorption spectra from 663, 645 and 652 nm by using a UV-1800 spectrophotometer and the amount of total chlorophyll, chlorophyll a and b determined as $\mu\text{g}/100\text{cm}^2$ with correction.

Analysis of Fatty Acid Composition

For detecting the total lipid values; the samples were homogenised and extracted in methanol/chloroform (2:1; v:v) according to Folch et al.

The fatty acid composition was analyzed by using a GC-FID (Gas Chromatography, Flame Ionization Detector). For this reason; aliquots of the extracted samples were taken. The lipid fraction was saponified with a 0.5 N methanolic NaOH solution. The fatty acids were methylated with $\text{BF}_3:\text{MeOH}$ and the resulting esters were analyzed.

Shimadzu GC (Shimadzu, GC-2014, Japan) was used for the determination of the composition of the fatty acids of the samples. GC solution software was used in order to control the system. A capillary column TRB-WaxOmega with dimensions of 30 m* 0.25 mm I.D*0.25 μm film thickness (Teknokroma, Spain) was used for the chromatography of the fatty acid methyl

esters. GC conditions were performed, such as injection: Split 1:10 at 260°C, 2.0 µL; carrier gas: nitrogen 0.8 mL min⁻¹; oven program: the initial temperature of 70 °C for 2 min, 4 °C min⁻¹ from 70 °C to 150 °C, hold at 150 °C for 10 min, 3 °C min⁻¹ from 150 °C to 180 °C, hold at 180 °C for 12 min, 2 °C min⁻¹ from 180 °C to 200 °C, hold at 200 °C for 15, 2 °C min⁻¹ from 200 °C to 220 °C, hold at 220 °C for 10 and then finally raised to 240 °C at the rate of 10 °C min⁻¹ and hold at 240 °C for 10 min.

The peak identification of the fatty acids in the analyzed samples was carried out by comparing it to the retention times and the spectra of known standards used a Supelco 37 Component FAMES (fatty acid methyl esters) Mix. The instrument was calibrated with 0.1, 0.5, 1.00, 2.50 and 5.00 mg/L concentrations using a standard solution of Supelco 37 Component FAMES (Folch et al., 1957).

The statistical analysis was conducted using the SAS.9.1.3. Portable Computer Pocket Program and controlled by a LSD test. Biplot used this for comment to fatty acid conclusions.

Results and Discussion

As it can be seen in Table 1, the highest fruit weights were obtained from Kcp3 level of irrigation and more fruit weight was obtained by increasing the level of irrigation (Demirel et al., 2012). With Proline applications at Kcp=0.5 the irrigation level increased the fruit weight whereas statistically the same fruit weight was obtained with the proline application at Kcp=1 and Kcp=1.5 level of irrigations.

As represented in Table 2, Kcp=1 and Kcp=1.5 level of irrigations had the same fruit number values which were more than Kcp=0.5 level of irrigation. Plants with Kcp=0.5 level of irrigation with proline application had statistically higher values than plants with no proline application in this irrigation level. Proline applications at Kcp=1 and Kcp=1.5 level of irrigation had no statistical difference regarding the fruit number. Ünükara et al. (2015) determined that a higher reduction of the amount of water decreased the fruit number whereas it remained unchanged or even increased at a proper reduction of water.

The yield per plant decreased from Kcp=1.5 level of irrigation to Kcp=1 and Kcp=0.5 level of irrigation and from major to minor respectively (Anjum et al., 2011) and the proline applications at Kcp=0.5 level of irrigation reduced the impact of water degradation on yield per plant (Table 3).

At Table 4, the contents of the total chlorophyll were expressed as chlorophyll a and chlorophyll b. The irrigation level enhanced the amount of the total chlorophylls. The proline application enhanced the total chlorophyll while this enhancement was more pronounced at Kcp=0.5 level of irrigation. It was reported that drought stress decreased the chlorophyll content (Oliveira Neto et al., 2009; Chegah et al., 2013; Moustakas et al., 2011).

A decline was determined at chlorophyll a contents with a lower level of irrigation. Proline applications caused a statistical difference at Kcp=0.5 and

Kcp=1 level of irrigations whereas no difference was determined at Kcp=1.5 level of irrigation (Table 5).

As shown in Table 6, Kcp=1.5 and Kcp=1 level of irrigations had similar and more chlorophyll b contents than Kcp=0.5 level of irrigation and proline applications increased the chlorophyll b content at Kcp=0.5 level of irrigation.

It was reported that exogenous proline positively affected yield and physiological parameters on salt (Öztekin, 2009) and drought stress (Ali et al., 2007).

The biplot comparison metot (Figure 1) was used to compare the effects of drought and proline applications and to identify differences on some fatty acid compositions of pepper. The biplot displays 76% of the information in the standardized data of the 3 irrigation levels and the proline applications for 4 fatty acids. Relative vector lengths of fatty acids didn't show an apparent difference, implying that all fatty acid factors equally contributed to total variation shown in the biplot graphic. Subjects that have higher palmitic, stearic and linoleic acid contents had lesser alfa linolenic acid contents as seen in biplot graphic (Figure 1), which shows a negative association between alfa linolenic acid and other fatty acids. The proline application to Kcp=0.5 elevated the alfa linolenic acid content as suggested by (Ali et al., 2013), and slightly decreased the stearic acid content, whereas the stearic acid continually decreased with higher levels of irrigations (Kcp=1.5). The alfa linolenic acid content increased and the stearic acid decreased in 3 (Kcp=1) and 5 (Kcp=1.5) compared to 1 (Kcp=0.5). These results were obtained by Junior et al., 2008; Hamrouni et al., 2001; Guerfel et al., 2008 on drought and Zhang et al., 2005 on salt stress. The alfa linolenic acid content decreased in 6 compared to 5 while 6 had the highest linolenic acid content among all treatments. The effect of the proline application was more obvious between treatments 1 and 2 compared to well watered treatments (Figure 1 and Table 7).

Table 1. *Effects of Different Levels of Irrigation and Proline Applications on the Fresh Fruit Weight*

	NP	PR	Kcp mean
Kcp1=0.5	48.67 D	52.08 C	50.37 C
Kcp2=1	67.71 B	68.05 B	67.88 B
Kcp3=1.5	75.26 A	75.47 A	75.36 A
P mean	63.88 B	65.20 A	

Shown are NP: No proline application, PR: Proline application; LSD values for P ≤0.01 PR×Kcp=1.9223; Kcp=2.6259; PR=0.9066

Table 2. *Effects of Different Levels of Irrigation and Proline Applications on the Fruit Number*

	NP	PR	Kcp mean
Kcp1=0.5	8.16 C	8.46 B	8.31 B
Kcp2=1	9.8 A	9.8 A	9.8 A
Kcp3=1.5	9.8 A	9.8 A	9.8 A
P mean	9.25 B	9.35 B	

Shown are NP: No proline application, PR: Proline application; LSD values for $P \leq 0.01$ $PR \times Kcp = 0.2485$; $Kcp = 0.3315$; $P \leq 0.05$ $PR = 0.0816$

Table 3. *Effects of Different Levels of Irrigation and Proline Applications on the Yield per Plant*

	NP	PR	Kcp mean
Kcp1=0.5	397.48 C	441.02 D	419.25 C
Kcp2=1	663.67 B	666.93 B	665.3 B
Kcp3=1.5	737.59 A	739.7 A	738.64 A
P mean	599.58 B	615.88 A	

Shown are NP: No proline application, PR: Proline application; LSD values for $P \leq 0.01$ $PR \times Kcp = 34.29$; $Kcp = 48.257$; $PR = 14.925$

Table 4. *Effects of Different Levels of Irrigation and Proline Applications on the Total Chlorophyll*

	NP	PR	Kcp mean
Kcp1=0.5	26.02 E	28.67 D	27.34 C
Kcp2=1	33.34 C	33.58 CB	33.46 B
Kcp3=1.5	35.04 AB	35.13 A	35.08 A
P mean	31.47 B	32.46 A	

Shown are NP: No proline application, PR: Proline application; LSD values for $P \leq 0.01$ $PR \times Kcp = 1.5289$; $Kcp = 1.4181$; $P \leq 0.05$ $PR = 0.7222$

Table 5. *Effects of Different Levels of Irrigation and Proline Applications on Chlorophyll a*

	NP	PR	Kcp mean
Kcp1=0.5	19.65 D	21.47 C	20.56 C
Kcp2=1	25.06 B	25.19 BA	25.12 B
Kcp3=1.5	26.44 A	26.45 A	26.45 A
P mean	23.71 B	24.37 A	

Shown are NP: No proline application, PR: Proline application; LSD values for $P \leq 0.01$ $PR \times Kcp = 1.2695$; $Kcp = 1.1622$; $P \leq 0.05$ $PR = 0.6035$

Table 6. Effects of Different Levels of Irrigation and Proline Applications on Chlorophyll b

	NP	PR	Kcp mean
Kcp1=0.5	6.38 C	7.21 B	6.80 B
Kcp2=1	8.3 A	8.42 A	8.36 A
Kcp3=1.5	8.61 A	8.69 A	8.65 A
P mean	7.76 B	8.11 A	

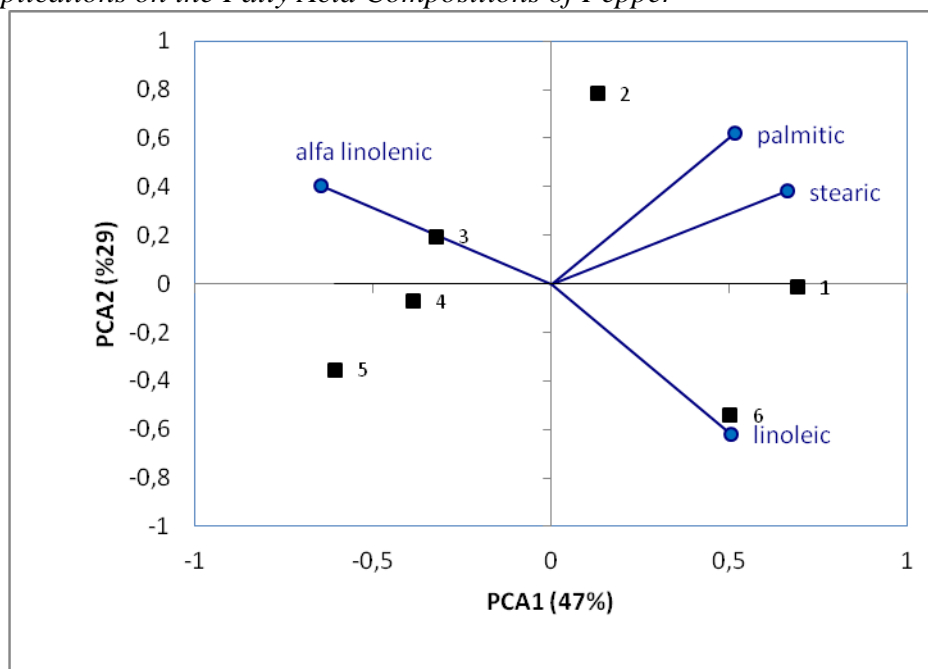
Shown are NP: No proline application, PR: Proline application; LSD values for $P \leq 0.01$ $PR \times Kcp = 0.3995$; $Kcp = 0.3246$; $PR = 0.3016$

Table 7. Effects of Different Levels of Irrigation and Proline Applications on the Fatty Acid Compositions of Pepper (%)

Subject	Linoleic	Alfa Linolenic	Palmitic	Stearic
1	39.75	19.28	20.87	3.78
2	38.81	25.68	22.68	3.51
3	38.83	25.92	20.13	3.39
4	39.50	26.51	19.30	3.40
5	38.93	23.83	19.49	2.91
6	42.21	22.03	20.93	3.39

Shown are 1: Kcp1=0.5, no proline applied; 2: Kcp1=0.5, proline applied; 3: Kcp2=1, no proline applied; 4: Kcp2=1, proline applied; 5: Kcp3=1.5, no proline applied; 6: Kcp3=1.5, proline applied

Figure 1. Biplot of Effects of Different Levels of Irrigation and Proline Applications on the Fatty Acid Compositions of Pepper



Shown are 1: Kcp1=0.5, no proline applied; 2: Kcp1=0.5, proline applied; 3: Kcp2=1, no proline applied; 4: Kcp2=1, proline applied; 5: Kcp3=1.5, no proline applied; 6: Kcp3=1.5, proline applied

Conclusions

According to the results; fruit weight, fruit number, yield per plant, total chlorophyll, chlorophyll a and b values were increased in Kcp=1 and Kcp=1.5 irrigation levels whereas decreased in Kcp=0.5. Fruit weight, fruit number, yield per plant, total chlorophyll, chlorophyll a and b values were increased by the proline applications at Kcp=0.5 irrigation level. This shows that, effects of proline application was more obvious in the deficient (Kcp=0.5) irrigation regime in terms of some (fresh fruit weight, yield, total chlorophyll, chlorophyll a and b) parameters. Kcp=1.5 level of irrigation with proline application had the highest fruit weight, yield and chlorophyll contents. Treatments with Kcp=1 and Kcp=1.5 level of irrigation and Kcp=0.5 level of irrigation with proline application had a higher alfa linolenic and lower stearic acid contents than Kcp=0.5 level of irrigation. The highest linoleic acid content found in proline applied Kcp=1.5 irrigation level while other treatments showed similar linoleic acid content.

References

- Ahire R. K., Kale A. A., Munjal S., V., Jamdagni B.M. 2005. Induced Water Stress Influencing Proline Accumulation, Protein Profiles and DNA Polymorphism In Chickpea Cultivars. *Indian Journal of Plant Physiology* 10 (3): 218–224.
- Ali, Q., Ashraf, M., Athar, H.U.R. 2007. Exogenously Applied Proline At Different Growth Stages Enhances Growth Of Two Maize Cultivars Grown Under Water Deficit Conditions. *Pak. J. Bot.*, 39(4): 1133-1144, 2007.
- Ali, Q., Anwar, F., Ashraf, M., Saari, N., Perveen, R. 2013. Ameliorating Effects of Exogenously Applied Proline on Seed Composition, Seed Oil Quality and Oil Antioxidant Activity of Maize (*Zea mays L.*) under Drought Stress. *International Journal of Molecular Sciences*, 2013, 14, 818-835; doi:10.3390/ijms14010818.
- Alvino, A., Centritto, M., and De Lorenzi., F. 1994. Photosynthesis response of sunlit and shade pepper (*Capsicum annuum*) leaves at different positions in the canopy under two water regimes. *Austral. J. Plant Physiol.* 21:377–391.
- Anjum, S.A., Farooq, M., Xie, X., Liu, X., Ijaz, M.F. 2011. Antioxidant defense system and proline accumulation enables hot pepper to perform better under drought. *Scientia Horticulturae* 140 (2012) 66-73.
- Ayers, R.S. and Westcot, D.W. 1989. Water quality for agriculture. Irr. Drainage Paper 29 rev. 1. FAO. Rome.
- Blum, A. 1986. Breeding Crop Varieties for Stress Environments. *Critical Reviews in Plant Sciences*, 2: 199-237.
- Chegah, S., Chehrazi, M., Albaji, M. 2013. Effects of drought stress on growth and development Frankenia plant (*Frankenia Leavis*). *Bulg. J. Agric. Sci.*, 19: 659-665.
- Dakhma, W.S., Zarrouk, M., Cherif, A. 1995. Effects of drought-stress on lipids in rape leaves. *Phytochemistry* 40:1383-1386.
- Demirel, K., Saçan, M., Genç, L. 2012. Yarı Kurak Koşullarda Farklı Sulama Düzeylerinin Salçalık Biberde (*Capsicum Annuum* cv. Kapija) Verim ve Kalite Parametreleri Üzerine Etkisi. [Effects of Different Irrigation Levels On Pepper

- (*Capsicum Annum* Cv. Kapija) Yield And Quality Parameters in Semi-Arid Conditions]. *Tekirdağ Ziraat Fakültesi Dergisi, Journal of Tekirdag Agricultural Faculty*, 9 (2): 7-15.
- Folch, J., Lees, M. & Sloane-Stanley, G.H. 1957. A simple method for the isolation and purification of total lipids from animal tissue, *Journal of Biological Chemistry*, 226 (1957) 497-509.
- Ghafoorunissa and Pangrekar, J. 1993. Vegetables as source of α -linolenic acid in Indian diets. *Food chemistry*, 47: 121–124.
- Gigon, A., Matos, A-R., Laffray, D., Zuily-Fodil, Y., Pham-Thi, A-T. 2004. Effect of drought stress on lipid metabolism in the leaves of *Arabidopsis thaliana* (Ecotype Columbia). *Ann Bot* 94:345-351.
- Gogos, U. and Smith, C. 2010. n-3 Omega fatty acids: a review of current knowledge. *Int. J. Food Sci. Technol.*, 45: 417–436. acid: A review. *Altern. Med. Rev.* 6(4): 367-382.
- Guerfel, M., O. Baccouri, D. Boujnah, and M. Zarrouk. 2008. Changes in lipid composition, water relations and gas exchange in leaves of two young ‘Chemlali’ and ‘Chetoui’ olive trees in response to water stress. *Plant Soil* 311:121–129.
- Hamrouni, I., H.B. Salah, B. Marzouka. 2001. Effects of water-deficit on lipids of safflower aerial parts. *Phytochemistry* 58:277–280.
- Holden, M. 1976. Chlorophyll in Chemistry and Biochemistry of Plant Pigments. Vol. 2 (T. W. Goodwin, Ed.). Academic Press, London pp: 1 – 37.
- Holub, B.J. 2002. Clinical nutrition: 4. Omega-3 fatty acids in cardiovascular care. *Can Med. Assoc. J. (JMAC)* 166 (5): 608 - 615.
- Huang, B., Zhong, D., Du, H., Wang, Z. 2011. Genotypic Variation in Fatty Acid Composition and Unsaturation Levels in Bermudagrass Associated with Leaf Dehydration Tolerance. *J. AMER. SOC. HORT. SCI.* 136(1):35–40. 2011.
- Junior, R.R.M., Oliveira, M.S.C., Baccache, M.A., Paula, F.M. 2008. Effects of water deficit and rehydration on the polar lipid and membranes resistance leaves of *Phaseolus vulgaris* L. cv. Pérola. *Braz. arch. biol. technol.* vol.51 no.2 Curitiba.
- Liu, L., Howe, P., Zhou, Y., Hocart C., Zhang R. 2002. Fatty acid profiles of leaves of nine edible wild plants: an Australian study. *Journal of Food Lipids*, 9: 65–71.
- Martinez, S., Curros, A., Bermudez, J., Carballo, J., Franco, I. 2006. Fatty acid profile of the fat from three pepper varieties (Arnoia, Fresno de la and Los Valles-Benavente). Effect of the ripening stage. *Grassas y Aceites*, Vol 57, No 4 (2006).
- Meiri, A. and Shalhevet, J. 1973. Crop growth under saline conditions, p. 277-290. In: B. Yaron, E. Danfors and Y. Vaadia (eds.). *Arid zone irrigation. Ecol. Studies* 5. Springer-Verlag, New York.
- Moustakas, M., Sperdoui, I., Kouna T., Antonopoulou, C.I., Therios, I. 2011. Exogenous proline induces soluble sugar accumulation and alleviates drought stress effects on photosystem II functioning of *Arabidopsis thaliana* leaves. *Plant Growth Regulation*, v.65, no.2, 2011 Nov, p.315 (11).
- Murphy, D.J. 1993. Structure, Function and Biogenesis of Storage Lipid Bodies and Oleosins in Plants, *Prog. Lipid. Res.* 32: 247–280.
- Oliveira Neto, C.F., Silva Lobato, A.K., Gonçalves-Vidigal, M.C., Lobo Da Costa, R.C., Santos Filho, B.G., Ruffeil Alves, G.A., Mello E Silva Maia, W.J., Rodrigues Cruz, F.J., Borges Neves, H.K., Santos Lopes, M.J. 2009. Carbon Compounds and Chlorophyll Contents in Sorghum Submitted to Water Deficit During Three Growth Stages. *Journal of Food, Agriculture & Environment* 7 (3-4) : 588- 593.

- Oztekin, G.B. 2009. Aşılı Domates Bitkilerinde Tuz Stresine Karşı Anaçların Etkisi (Doktora Tezi) [Response Of Tomato Rootstocks To Salinity Stress (PhD Thesis)]. Ege Üniversitesi, İzmir. 43-44.
- Pereira, C., Li, D., Sinclair, A.J. 2001. The α -linolenic acid content of green vegetables commonly available in Australia. *International Journal of Vitamin and Nutrition Research*, 71: 223–228.
- Perez-Galvez, A., Garrido-Fernandez, J., Minguez-Mosquera, I., Lozano-Ruiz, M., Montero-de-Espinosa, V. 1999. Fatty Acid Composition of Two New Pepper Varieties (*Capsicum annuum* L. cv. *Jaranda* and *Jariza*). Effect of Drying Process and Nutritional Aspects. Paper no. J8983 in *JAOCs* 76, no. 2, 205–208 (February 1999).
- Rhoades, J.D., Kandiah, A. and Mashali, A.M. 1992. The use of saline waters for crop production. *Irr. Drainage Paper* 48. FAO, Rome.
- Unlukara, A., Kurunc, A., Cemek, B. 2015. Green Long Pepper Growth under Different Saline and Water Regime Conditions and Usability of Water Consumption in Plant Salt Tolerance. *Journal Of Agricultural Sciences*, 21: 167-176.
- Vieira da Silva, J., Naylor, A.W., Kramer, J. 1974. Some ultrastructural and enzymatic effects of water stress in cotton (*Gossypium* L.) leaves. *Proceedings of the National Academy of Sciences of the USA* 71: 3243–3247.
- Vural, H., Esiyok, D. ve Duman, I. 2000. Kültür Sebzeleri [Cultured Vegetables], Ege Üniversitesi, Ziraat Fakültesi Bahçe Bitkileri Bölümü, Bornova-İZMİR, ISBN:975-97190-0-2, 293-306.
- Yildirim, O. 1996. Sulama Sistemleri 2 [Irrigation Systems 2]. Tarımsal Yapılar ve Sulama Bölümü. Ziraat Fakültesi. Ankara Üniversitesi. Yayın No: 1449, Ankara. 354 p.
- Zhang, M., Barg, R., Yin, M., Gueta-Dahan, Y., Leikin-Frenkel, A., Salts, Y., Shabtai, S., Ben Hayyim, G. 2005. Modulated fatty acid desaturation via overexpression of two distinct ω -3 desaturases differentially alters tolerance to various abiotic stresses in transgenic tobacco cells and plants. *Plant J* 44:361-371.